



Short Research Communication

Characterization of the Asian Citrus Psyllid Transcriptome

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Abstract

The Asian citrus psyllid, *Diaphorina citri* Kuwayama (Hemiptera: Psyllidae) is a vector for the causative agents of Huanglongbing, which threatens citrus production worldwide. This study reports and discusses the first *D. citri* transcriptomes, encompassing the three main life stages of *D. citri*, egg, nymph and adult. The transcriptomes were annotated using Gene Ontology (GO) and insecticide-related genes within each life stage were identified to aid the development of future *D. citri* insecticides. Transcriptome assemblies and other sequence data are available for download at the International Asian Citrus Psyllid Genome Consortium website [<http://psyllid.org/download>] and at NCBI [<http://www.ncbi.nlm.nih.gov/bioproject/29447>].

Key words: Asian Citrus Psyllid, *Diaphorina citri* Kuwayama

Results and Discussion

General Characteristics of the *D. citri* Transcriptomes

In total, 46,927,970 reads of 75 bp, 39,830,860 reads of 110 bp, and 50,248,212 reads of 100 bp were generated from the egg, nymph, and adult tissues, respectively, comprising 12.9 Gb of sequence, and

were used to construct three *de novo* stage-specific transcriptomes (Table 1). The GC content of the transcriptomes was highly similar, ranging from 42.10% to 44.81%. Moreover, 99 to 100% of core eukaryotic genes had a detectable homolog in the stage-specific assemblies and between 84.3 and 88.2% of core genes had a detectable homolog whose alignment covered

>80% of the core gene, indicating the transcriptomes were fairly complete.

Functional Annotation and Characterization of *Diaphorina citri* Transcripts

To review the putative functions of the *D. citri* transcripts and validate the completeness of the transcriptomes, GO analysis was performed against the three *D. citri* stages, plus four related organisms with complete transcriptome information. Of the total number of representative transcripts present within the egg, nymph and adult stages, 45.20% (26,546), 46.29% (26,651), and 40.00% (21,218) were assigned GO terms, respectively (Figure 1). Importantly, all four related organisms displayed this pattern, suggesting the completeness of the three stage transcriptomes.

Identification of *Diaphorina citri* Insecticide-Related Genes

Insecticides are a pivotal component in controlling *D. citri* populations throughout the world, and therefore, it is essential to develop potent insecticides against a wide variety of molecular *D. citri* targets [1]. Insecticide targets and genes involved in detoxification and resistance were identified in the egg, nymph, and adult stages (Figure 2A). The largest and most complex group was 'Juvenile Hormone Metabolism.' Putative *D. citri* homologs of the major gene products involved in juvenile hormone metabolism exist within each life stage and exhibit a high degree of similarity to those genes of closely related organisms (Figure 2B and Table 2).

Table 1. Statistics for *Diaphorina citri* transcriptome assemblies.

Assembly	Stage	Total Number of Transcripts	Mean Contig Length	GC%	Core Eukaryotic Genes with Homologs	Core Eukaryotic Genes with Homologs (>80% coverage)	% of Transcripts Confirmed in Draft Genome Assembly	Putative Gene Fusions
Complete	Egg	76,550	852	42.1	458/458 (100%)	400/458 (87.3%)	81.6%	196 (0.256%)
-	Nymph	69,233	756	44.81	456/458 (99.56%)	404/458 (88.2%)	81.0%	146 (0.211%)
-	Adult	62,450	691	42.96	458/458 (100%)	386/458 (84.3%)	81.0%	158 (0.253%)
Representative*	Egg	58,814	719	41.83	458/458 (100%)	400/458 (87.3%)	84.2%	163 (0.278%)
-	Nymph	57,635	632	44.27	456/458 (99.56%)	403/458 (88.0%)	79.0%	123 (0.214%)
-	Adult	53,117	595	42.67	458/458 (100%)	386/458 (84.3%)	88.7%	147 (0.277%)

Files containing the assembled transcripts for each of the *D. citri* life stages were analyzed using custom Perl scripts, and the total number of transcripts, mean contig length, %GC, number of core eukaryotic genes, % transcripts detectable in the genome assembly and number of putative gene fusions were determined. (See Methods section for details about how each was calculated.)

Table 2. Juvenile hormone related genes identified in the *Diaphorina citri* transcriptome.

Gene Name	NCBI Gene ID	FlyBase ID	Query ID	Query Length (bp)	Subject ID	Subject Species	E-Value
Allatostatin	42947	FBgn0015591	diaci_nymph_66660000039047	692	NP_001037036.1	<i>Bombyx mori</i>	3e-07
Allatostatin Receptor	44126	FBgn0028961	diaci_nymph_66660000026007	1547	NP_524700.1	<i>Drosophila melanogaster</i>	0
Broad	44505	FBgn0000210	diaci_nymph_66660000033288	578	NP_726750.1	<i>Drosophila melanogaster</i>	0
Chd64	38490	FBgn0035499	diaci_nymph_66660000018386	2227	AAF47840.2	<i>Drosophila melanogaster</i>	1e-136
Cytosolic Juvenile Hormone Binding Protein	733092	-	diaci_nymph_66660000009996	768	NP_001037668.1	<i>Bombyx mori</i>	4e-142
FKBP39	41860	FBgn0013269	diaci_nymph_66660000005954	2150	CAA86996.1	<i>Drosophila melanogaster</i>	0
Hexamerin	660274	-	diaci_nymph_66660000015491	2322	NP_001164358.1	<i>Tribolium castaneum</i>	0
Juvenile Hormone Acid Methyltransferase	34977	FBgn0028841	diaci_egg_55550000038216	1502	EFA02917.1	<i>Tribolium castaneum</i>	3e-126
Juvenile Hormone Epoxide Hydrolase	251984	FBgn0010053	diaci_nymph_66660000003152	1779	NP_001161902.1	<i>Tribolium castaneum</i>	2e-49
Juvenile Hormone Esterase	36780	FBgn0010052	diaci_adult_77770000038704	271	NP_001180223.1	<i>Tribolium castaneum</i>	9e-84
Juvenile Hormone Esterase Binding Protein	37996	FBgn0035088	diaci_nymph_66660000032724	733	NP_611989.1	<i>Drosophila melanogaster</i>	2e-18
Methoprene-Tolerant	32114	FBgn0002723	diaci_egg_55550000026684	389	ABR25244.1	<i>Tribolium castaneum</i>	8e-39
Retinoid X Receptor	31165	FBgn0003964	diaci_nymph_66660000021934	2841	AAF45707.1	<i>Drosophila melanogaster</i>	5e-93

Putative juvenile hormone related genes were identified using BLASTX to compare the representative *D. citri* transcripts from the egg, nymph and adult life stages to a custom BLAST database containing those genes of interest from closely related organisms. The top BLAST hits for the juvenile hormone related genes are shown.

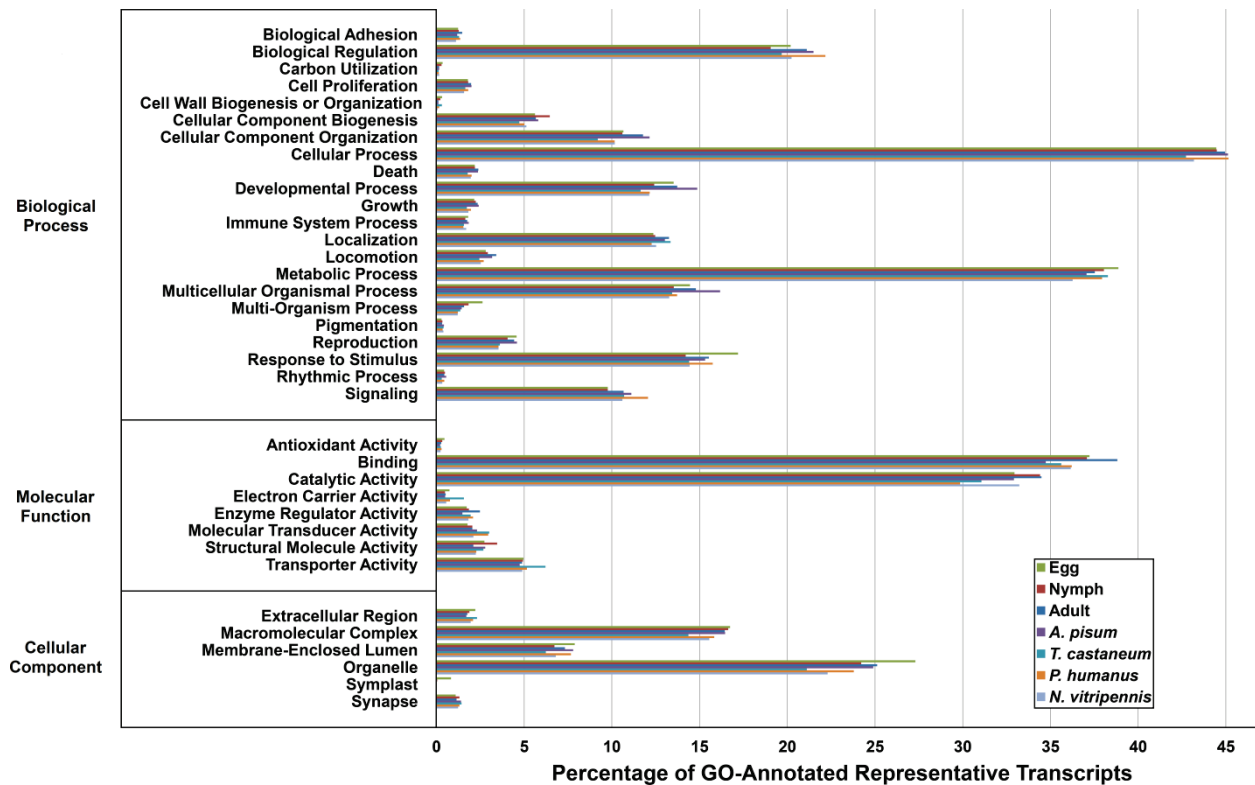


Figure 1. Gene Ontology (GO) analysis of the representative transcripts present within the three life stages of *Diaphorina citri*. GO-terms were assigned to the representative transcripts of the three *D. citri* life stages and *Acyrtosiphon pisum*, *Tribolium castaneum*, *Pediculus humanus* and *Nasonia vitripennis*, the four organisms that exhibited the greatest number of top BLAST hits, using B2G4Pipe and then sorted into groups within three independent compartments using Blast2GO. The ‘Cellular Process’, ‘Binding’ and ‘Organelle’ groups contain the greatest number of representative transcripts within the ‘Biological Processes’, ‘Molecular Function’ and ‘Cellular Component’ domains, respectively. The representative transcriptomes of the four organisms and the egg, nymph and adult stages exhibited similar patterns, and thus indicates the completeness of the stage transcriptomes.

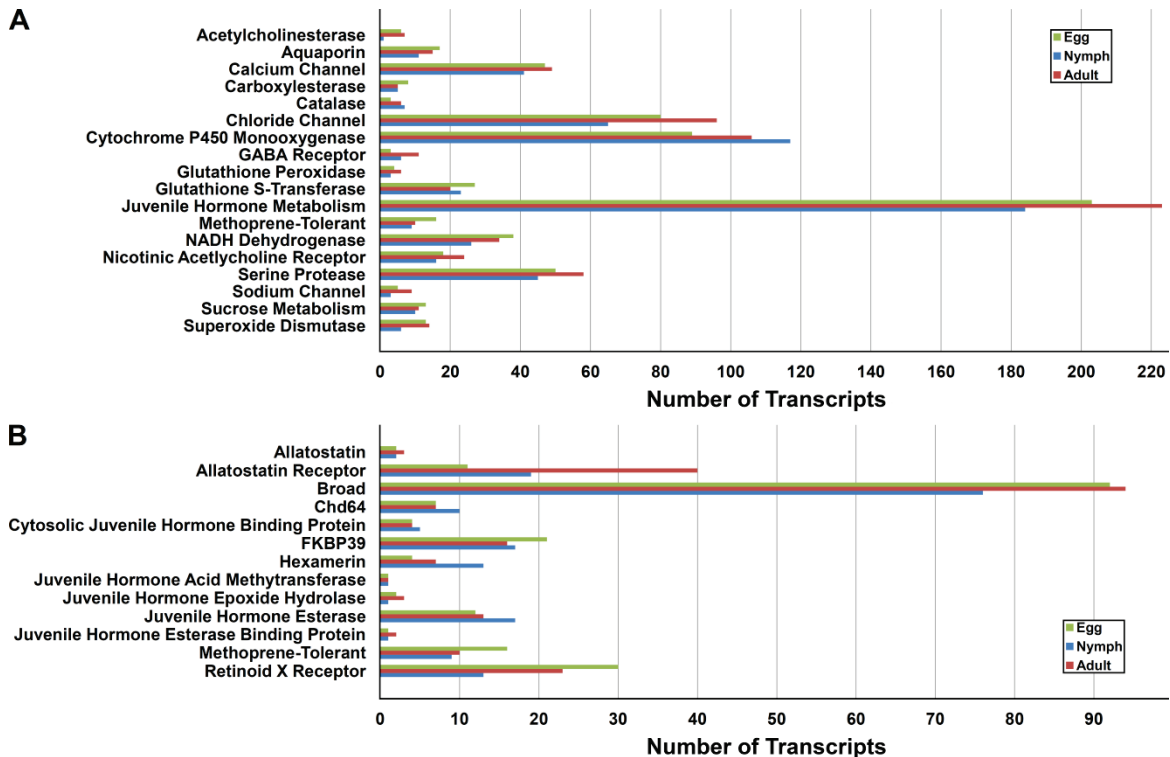


Figure 2. Genes related to insecticide targets, detoxification, and resistance within each of the three life stages of *Diaphorina citri*. Identification of putative *D. citri* insecticide-related genes was accomplished by comparing the representative transcripts from the three *D. citri* life stages to a custom protein database containing homologous genes of interest from closely related organisms using BLASTX. (A) *D. citri* predicted gene products of putative insecticide targets and predicted gene products putatively involved in detoxification and resistance were identified in all three life stages. (B) *D. citri* homologs of the major gene products involved in insect juvenile hormone metabolism are present within each life stage.

Methods

Growth, Sample Preparation, and Sequence Generation

Psyllids were field collected from citrus groves near USDA-ARS research station, 2001 South Rock Road, Fort Pierce, FL, 34945-3030; no specific permits were required. Psyllids were reared in 2' x 2' cages, temperature maintained at 26° C, lighting was dependent on natural day length throughout the year, and fed on *Murraya paniculata* for two years then *Citrus macrophylla* for the last two years. Tissue from one to four day old eggs, 3rd and 4th instar nymphs, and one day to one month old adults of mixed genders were collected, processed, and held at -80°C until RNA isolation.

Total RNA was isolated from whole egg, nymph, and adult tissues using Qiagen's RNeasy Mini Kit, per the manufacturer's instructions. The mRNA was purified using poly-T oligo-attached magnetic beads and converted into cDNA with random primers using the mRNA sequencing preparation kit from Illumina (part number 1004898). The cDNA was sequenced using an Illumina GAIIx sequencing system.

Transcriptome Assembly and Characterization

Velvet (v1.0.19) (k-mer of 47) and Oases (v0.1.19) were used to generate three stage-specific transcriptomes using reads from the egg, nymph or adult tissues, respectively [2,3]. Contigs with adapter sequence contamination were removed by alignment to Illumina adapter sequences using BWA [4].

The GC content and mean transcript length, were calculated using a custom Perl script to check completeness, transcriptomes were aligned with TBLASTN to sequences representing 458 core eukaryotic genes [5], and percent of core genes with at least one KOG family member that aligned to a transcript with an *E*-value < 1e-6 (optionally with alignment that covered 80% of KOG family member) was measured. To detect gene fusions, transcripts were aligned using BLASTN to 16,172 gene predictions produced using MAKER [134], and considered gene fusions if 1) the alignments between the transcript and two different MAKER gene predictions had an *E*-value < 1e-6 and percent identity >95%, 2) coordinates of the two alignments on the transcript overlapped by <10 nucleotides, 3) the two alignments together covered >95% of transcript sequence and 4) no alignment existed between transcript and MAKER model with an *E*-value <1e-6 and percent identity >95% that covered >95% of the transcript.

Generation of Representative Transcriptome Transcripts

To generate transcriptomes containing only unique transcripts for downstream analysis, by removing all repetitive, identical and near-identical transcripts, CD-HIT-EST was used with a sequence identity cut-off of 99% [6,7]. A loss of 23.17% (17,736), 16.75% (11,598), and 14.94% (9,333) of the total transcripts from the egg, nymph, and adult stages, respectively, was observed. Also, in order to compare *D. citri* to other previously sequenced Insecta spp., all NCBI RefSeq nucleotide records (retrieved August 12, 2012) were collected for *Acyrtosiphon pisum*, *Tribolium castaneum*, *Pediculus humanus* and *Nasonia vitripennis* totaling 17,675; 10,417; 10,775 and 12,927 sequences, respectively. In order to generate representative transcriptomes for each of these species, near-identical transcripts were removed using CD-HIT-EST using a cut-off value of 99% [6,7]. This resulted in a 4.64% (821), 4.09% (426), 0.47% (51) and 2.07% (267) loss of the total number of transcripts for *A. pisum*, *T. castaneum*, *P. humanus* and *N. vitripennis*, respectively.

Gene Ontology

To infer the putative function of the *D. citri* transcripts, BLASTX and an *E*-value threshold of $\leq 1e-5$ was used to scan the representative transcriptomes against the NCBI non-redundant (nr) database (retrieved April 23, 2012). GO terms were assigned and were then categorized using the programs B2G4Pipe (version 2.5.0) and Blast2GO (version 2.5.1) using default settings, respectively, and the b2g_may12 GO database [8].

Identification of Insecticide-Related Genes

Insecticide-related genes present within the different life stages of *D. citri* were identified by using a combination of BLASTN, TBLAST, and BLASTX and an *E*-value threshold of $\leq 1e-5$ to compare the representative transcriptomes to a custom BLAST database containing insecticide-related genes from *A. pisum*, *Apis mellifera*, *Bombyx mori*, *Drosophila melanogaster*, *N. vitripennis*, *P. humanus*, and *T. castaneum* (retrieved from NCBI on July 30, 2012).

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Competing Interests

The authors have declared that no competing interests exist.

References

1. Hall, DG, Ammar ED, Richardson ML, and Halbert SE. Asian citrus psyllid, *Diaphorina citri* (Hemiptera: Psyllidae), vector of citrus huanglongbing disease. *Entomologia Experimentalis et Applicata* 2012; 146: 207-223.
2. Schulz MH, Zerbino DR, Vingron M, Birney E. Oases: robust de novo RNA-seq assembly across the dynamic range of expression levels. *Bioinformatics* 2012; 28: 1086-1092.
3. Zerbino DR, Birney E. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. *Genome Res* 2008; 18: 821-829.
4. Li H, Durbin R. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics* 2009; 25: 1754-1760.
5. Parra G, Bradnam K, Korf I. CEGMA: a pipeline to accurately annotate core genes in eukaryotic genomes. *Bioinformatics* 2007; 23: 1061-1067.
6. Li W, Godzik A. Cd-hit: a fast program for clustering and comparing large sets of protein or nucleotide sequences. *Bioinformatics* 2006; 22: 1658-1659.
7. Limin Fu, Beifang Niu, Zhengwei Zhu, Sitao Wu and Weizhong Li. CD-HIT: accelerated for clustering the next generation sequencing data. *Bioinformatics* 2012; 28 (23): 3150-3152. doi: 10.1093/bioinformatics/bts565.
8. Conesa A, Gotz S, Garcia-Gomez JM, Terol J, Talon M, et al. Blast2GO: a universal tool for annotation, visualization and analysis in functional genomics research. *Bioinformatics* 2005; 21: 3674-3676.